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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.		
10/617,734	07/14/2003	Gregory Gregoriadis	G0365.0365/P0365	3606		
	7590 01/09/200° HAPIRO MORIN & O	EXAM	EXAMINER			
Edward A. Meilman 41st Floor 1177 Avenue of the Americas New York, NY 10036-2714			SCHNIZER, I	SCHNIZER, RICHARD A		
			ART UNIT	PAPER NUMBER		
			1635			
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			01/09/2007	PAPER		

Please find below and/or attached an Office communication concerning this application or proceeding.

Advisory Action Before the Filing of an Appeal Brief

Application No.	Applicant(s)	
10/617,734	GREGORIADIS, GREGORY	
Examiner	Art Unit	
Richard Schnizer, Ph. D.	1635	

	Richard Schnizer, Ph. D.	1635	
The MAILING DATE of this communication appe	ears on the cover sheet with the o	correspondence add	ress
THE REPLY FILED <u>13 December 2006</u> FAILS TO PLACE THI	S APPLICATION IN CONDITION F	OR ALLOWANCE.	
1. The reply was filed after a final rejection, but prior to or of this application, applicant must timely file one of the follo places the application in condition for allowance; (2) a Notal Request for Continued Examination (RCE) in compliant time periods:	n the same day as filing a Notice of wing replies: (1) an amendment, aff otice of Appeal (with appeal fee) in o	Appeal. To avoid aba fidavit, or other evider compliance with 37 Cl	ce, which FR 41.31; or (3)
a) \square The period for reply expires $\underline{6}$ months from the mailing date	e of the final rejection.		
b) The period for reply expires on: (1) the mailing date of this a no event, however, will the statutory period for reply expire Examiner Note: If box 1 is checked, check either box (a) or TWO MONTHS OF THE FINAL REJECTION. See MPEP 7	later than SIX MONTHS from the mailin (b). ONLY CHECK BOX (b) WHEN THE 706.07(f).	g date of the final rejecti E FIRST REPLY WAS F	on. ILED WITHIN
Extensions of time may be obtained under 37 CFR 1.136(a). The date have been filed is the date for purposes of determining the period of exunder 37 CFR 1.17(a) is calculated from: (1) the expiration date of the set forth in (b) above, if checked. Any reply received by the Office late may reduce any earned patent term adjustment. See 37 CFR 1.704(b) NOTICE OF APPEAL	ktension and the corresponding amount shortened statutory period for reply orig or than three months after the mailing da	of the fee. The appropri inally set in the final Offi	ate extension fee ce action; or (2) as
 The Notice of Appeal was filed on <u>13 December 2006</u>. A of the date of filing the Notice of Appeal (37 CFR 41.37(a appeal. Since a Notice of Appeal has been filed, any rep AMENDMENTS 	a)), or any extension thereof (37 CF	R 41.37(e)), to avoid	dismissal of the
3. The proposed amendment(s) filed after a final rejection,	but prior to the date of filing a brief	, will <u>not</u> be entered b	ecause
 (a) They raise new issues that would require further co (b) They raise the issue of new matter (see NOTE below) (c) They are not deemed to place the application in beautiful appeal; and/or 	onsideration and/or search (see NO ow); etter form for appeal by materially re	TE below);	
(d) They present additional claims without canceling a NOTE: (See 37 CFR 1.116 and 41.33(a))		ected ciaims.	
4. The amendments are not in compliance with 37 CFR 1.15. Applicant's reply has overcome the following rejection(s	21. See attached Notice of Non-Co): See Continuation Sheet.	·	•
 Newly proposed or amended claim(s) would be a non-allowable claim(s). 	illowable if submitted in a separate,	timely filed amendme	nt canceling the
 7. For purposes of appeal, the proposed amendment(s): a) how the new or amended claims would be rejected is proof. The status of the claim(s) is (or will be) as follows: Claim(s) allowed: 		II be entered and an e	explanation of
Claim(s) objected to:			
Claim(s) rejected: <u>1,3,6-20,22,25-31 and 34-36</u> . Claim(s) withdrawn from consideration:			
AFFIDAVIT OR OTHER EVIDENCE			
 The affidavit or other evidence filed after a final action, be because applicant failed to provide a showing of good ar was not earlier presented. See 37 CFR 1.116(e). 			
9. The affidavit or other evidence filed after the date of filing entered because the affidavit or other evidence failed to showing a good and sufficient reasons why it is necessa	overcome all rejections under appe	al and/or appellant fai	ls to provide a
10. The affidavit or other evidence is entered. An explanation REQUEST FOR RECONSIDERATION/OTHER	on of the status of the claims after e	ntry is below or attact	ned.
 The request for reconsideration has been considered be See attached. 	ut does NOT place the application i	n condition for allowar	nce because:
12. ☐ Note the attached Information Disclosure Statement(s).13. ☐ Other:	(PTO/SB/08) Paper No(s)		

Continuation of 5. Applicant's reply has overcome the following rejection(s): The rejection of claim 17-20, 22, and 25-32 under 35 USC 112, second paragraph.

Response to Arguments

Applicant's arguments filed 12/15/06 have been fully considered but they are not persuasive for the reasons of record in the Action mailed 6/13/06. Applicant reviews the Felgner reference at pages 10-11 of the response, and concludes that there is no teaching or suggestion in Felgner of cationic liposomes in which polynucleotides encoding immunogens are entrapped in the intravesicular space. The Examiner disagrees. The reference need not disclose applicant's invention in one sentence or even in one paragraph. The reference must be considered as a whole to determine what it fairly teaches. Felgner clearly states that the invention embraces delivery of nucleic acids that express immunogens in the paragraph bridging columns 7 and 8. Felgner also clearly indicates that therapeutic agents and "biologically active agents" include polynucleotides that can express proteins (see e.g. column 7, lines 49-56 and column 8, lines 60-65). Further, Felgner clearly teaches that the lipids of the invention may be used to form liposomes, and that the liposomes may encapsulate bioactive agents. See column 15, lines 7-25. There is no reason at all to assume that these active agents do not include nucleic acids encoding immunogens.

Applicant also asserts that Felgner fails to teach any in vivo results, noting that all experiments in Felgner were carried out in cell lines in vitro. However Applicant does not set forth any analysis that would lead one to the conclusion that the disclosure of Felgner was not enabling for the induction of an immune response. Furthermore, the Weiner and Liu references provide ample teachings to support this enablement.

Page 3

Art Unit: 1635

Applicant argues at page 12 of the response that the Kirby reference fails to overcome the alleged deficiencies of Felgner because Kirby does not teach DNA vaccines. This is unpersuasive because Kirby was not relied upon to teach DNA vaccines, Kirby was relied upon to teach a dehydration-rehydration method of encapsulating solutes such as DNA into liposomes that is a simple method which provides excellent encapsulation yields while using mild conditions. The method also results in a greater proportion of oligo- and multilamellar vesicles which decrease the rate of loss of entrapped solutes (see paragraph bridging pages 982, and 983) and would be expected to exclude nucleases with greater success than unilamellar vesicles, thereby increasing the stability of the encapsulated nucleic acid.

Applicant asserts that the best argument for combining Kirby with Felgner is that it would have been obvious to try the combination. In support, Applicant argues that Kirby does not teach DNA vaccines, taught entrapment of E. coli genomic DNA, did not show functionality of entrapped DNA, and did not teach in vivo delivery of DNA. This is unpersuasive because Weiner and Liu each provide working examples of DNA vaccines, and Weiner suggested use of liposomes to deliver these vaccines. There is no reason of record to believe that entrapment of nucleic acids in liposomes would eliminate DNA expression construct activity in vivo. On the contrary, one of ordinary skill in the art would reasonably expect to decrease the susceptibility of the entrapped nucleic acids to nucleases, thereby increasing the stability of the encapsulated nucleic acids.

Applicant submits at page 12 that the Office's contention that one would have been motivated to use the method of Kirby because it provided excellent encapsulation yields is a hindsight justification which is not supported by either reference. This is unpersuasive because the method of Kirby resulted in a DNA encapsulation efficiency of 72% +/- 8.5%. See Table 1. This is objective evidence of excellent encapsulation yield. Furthermore, the method of Kirby has the advantages of being simple and mild. See title, page 979; column 1, lines 5-8 of second paragraph; Table I on page 980., and page 982, column 2, lines 1-3 of second full paragraph. Thus one would clearly have been motivated to use it to encapsulate nucleic acids for preparation of DNA vaccines.

Applicants arguments at page 12 indicating that there is nothing in the cited references regarding generating a cell based and humoral immune response is unpersuasive. The claimed methods steps are obvious for the reasons set forth in the rejection, and the nature of the produced immune response is considered to be an inherent result of the steps.

At page 13, Applicant asserts that the combination of references is improper because the cationic lipids of Felgner are used to form complexes with nucleic acids in which the nucleic acid is ionically bound to a preformed liposome, and not entrapped in the intravesicular space. This is unpersuasive for two reasons. First, Felgner does fairly teach a method of entrapping nucleic acids into the intravesicular space of liposomes. See column 15, lines 7-20 which clearly teaches encapsulation of bioactive agents into the lumen of liposomes. It is clear from the disclosure of Felgner as a whole that nucleic acids are considered to be bioactive agents, see e.g. abstract column 1,

lines 12-18, column 2, lines 7-11, and column 4, lines 10-14. Second, it would have been obvious to encapsulate the nucleic acids within the intravesicular space in order to protect the nucleic acids from nucleases, thereby increasing the stability of the nucleic acids in vivo. Applicant argues at page 14 that any basis for asserting that the loss of solutes from the multilamellar vesicles of Kirby would correlate with increased protection from nucleases, relative to that provided by unilamellar liposomes, lacks support in fact or logic. In support, Applicant argues that there is no disclosure that nucleases attack nucleic acids. This is unpersuasive because it is clear to one of ordinary skill in the art from the name "nucleases" that the activity of these enzymes is to attack and degrade nucleic acids. Applicant states that there is no basis for asserting that nucleases diffuse into liposomes, and notes that the carboxyfluorescein observed by Kirby to diffuse through liposomal membranes is very different from nucleases. This is unpersuasive. Kirby indicates at page 982, column 1, lines 4-8 that multilamellar liposomes, rather than unilamellar liposomes, should afford maximum protection against the effects of enzymes. Thus it is clear that those of ordinary skill in the art, aware of Kirby would have thought that multilamellar liposomes provided more protection against enzymes.

For these reasons the rejections are maintained.

including nucleases, than would unilamellar liposomes.

RICHARD SCHNIZER, PH.D. PRIMARY EXAMINER

Page 5